





Chemical synthesis of glycopeptide related to *T. cruzi* and cancer mucins

Vanessa Leiria Campo, Ivone Carvalho, Marcelo Dias Baruffi

Faculdade de Ciências Farmacêuticas de Ribeirão Preto - USP. Av. do Café S/N, CEP 14040-903, Ribeirão Preto - SP, Brazil. vlcampo@fcfrp.usp.br

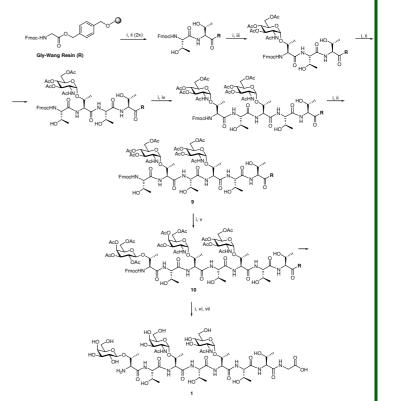
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INTRODUCTION

Mucins are highly O-glycosylated (about 60% carbohydrate by weight), glycoproteins that are rich in serine (Ser) and threonine (Thr) repeating units, whose external oligosaccharides are linked to the protein via sugar units of α -N-acetyl-glucosamine (GlcNAc), in T. *cruzi*, or α -*N*-acetylgalactosamine (GalNAc), in mammals.¹ In *T. cruzi*, these structures helps the parasite to interact with the infected host, while in vertebrates the functions of mucins range from being protective barriers to providing lubrication for epithelial cells. In cancer-related mucins, however, altered glycosylation is a common feature, being verified the presence of mono- and disaccharide structures known as T_{N} and TFantigens, respectively.² Therefore, this work presents the synthesis of the glycopeptide NH₂[BGal]-(Thr)₂- $[\alpha GalNAc]$ - $(Thr)_2$ - $[\alpha GlcNAc]$ - $(Thr)_3 GlvOH$ 1, related to T. cruzi and cancer mucins, which can be useful for development of new therapeutics, like vaccines, against infectious and tumoral processes.

RESULTS AND DISCUSSION

The synthesis of glycopeptide 1 involved the previous preparation of the glycosyl-amino acids αGlcNAc-ThrOH 2 (22%), αGalNAc-ThrOH 3 (54%), and ßGal-ThrOH 4 (33%) by glycosylation reactions between the corresponding sugars α GlcNAcCl 5, α GalN₃Cl 6 and α GalBr 7 with the amino acid FmocThrBn 8, followed by reductive acetylation reaction (compound 6) and removal of the benzyl ester groups by standard hydrogenation (10% Pd-C/ H_2).³ Subsequently, the synthesis of glycopeptide 1 in solid-phase was carried out in the presence of the benzotriazol-1-yloxytriscoupling reactants pyrrolidino phosphonium hexafluorophosphate (PyBOP) and 1-hydroxybenzotriazole (HOBt), and the base diisopropylethylamine (DIPEA) in DMF,³ as outlined in Scheme 1. After cleavage from the resin with aqueous TFA and removal of the sugar acetate protecting groups with catalytic NaOMe in MeOH, the glycopeptide 1 was purified by reverse-phase HPLC, being obtained in 20% overall yield. The structure of compound 1 was confirmed by NMR spectroscopy, whereas ESI-MS analysis is being realized.



Scheme 1. Solid-phase synthesis of glycopeptide 1. i. 20% piperidine-DMF ; ii. FmocThrOH, PyBOP, HOBt, DIPEA , 3h; iii, iv, and v. α GlcNAc-ThrOH 2, α GalNAc-ThrOH 3 or β Gal-ThrOH 4, PyBOP, HOBt, DIPEA, 24 h; vi. TFA 80%, vii. NaOMe, MeOH.

CONCLUSION

The glycopeptide **1** was obtained in reasonable yield by combination of *in solution* and solid phase methods, and will be further attached to a carrier protein (BSA or KLH) in order to be evaluated as potential anti-parasite and anti-cancer vaccine.

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CAPES and FAPESP

REFERENCES

- ¹ Buscaglia, C. A.; Campo, V. A.; Frasch, A. C. C.; DiNoia, J. M. *Nat. Rev. Microbiol.* **2006**, *4*, 229.
- Brocke, C.; Kunz, H. Bioorg. Med. Chem. 2002, 10, 3085.

³ Campo, V. L.; Carvalho, I.; Allman, S.; Davis, B. G.; Field, R. A. *Org. Biomol. Chem.* **2007**, *5*, 2645.

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